

Two New Iridoid Alkaloids, Daphmacropodosidines A and B, from *Daphniphyllum macropodum*

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Two new iridoid alkaloids, daphmacropodosidines A (**1**) and B (**2**), together with two known ones, daphcalycinosidine B and caldaphnidine F, were isolated from the fruits of *Daphniphyllum macropodum*. Their structures were established by spectral methods, especially 2D NMR spectra (¹H,¹H-COSY, HMQC, HMBC, and NOESY), as well as chemical means.

1. Introduction. – *Daphniphyllum* alkaloids with highly complex polycyclic structures are the secondary metabolites elaborated by plants of the genus *Daphniphyllum* [1]. Radioactive tracer experiments revealed that *Daphniphyllum* alkaloids were generated from six molecules of mevalonic acid *via* a squalene-like intermediate [2]. Heathcock and co-workers have done an outstanding work on a biomimetic total synthesis of several *Daphniphyllum* alkaloids [3]. In recent years, more than 60 new *Daphniphyllum* alkaloids were reported by Kobayashi, Bodo, Yue, Hao, and their co-workers [4–7]. However, only six iridoid alkaloids have been elucidated since the first one was reported by Kobayashi in 2003 [5][6c][8][9]. In our continuous research on structurally and biogenetically interesting *Daphniphyllum* alkaloids, two new iridoid alkaloids, daphmacropodosidines A (**1**) and B (**2**) (*Fig. 1*), together with two known ones, daphcalycinosidine B [5] and caldaphnidine F [6c], were isolated from the fruits of *Daphniphyllum macropodum*.

2. Results and Discussion. – Daphmacropodosidine A (**1**) was obtained as optically active, colorless crystals. Its molecular formula was established as C₄₀H₅₇NO₁₃ by HR-ESI-MS ([*M* + H]⁺ at *m/z* 760.3910), indicating the presence of 13 degrees of unsaturation. The IR spectrum of **1** exhibited absorptions for OH (3422 cm⁻¹), C=O (1729 and 1643 cm⁻¹), and C=C functions. The UV absorption at 307.2 nm suggested the presence of a conjugated system.

Hydrolysis of **1** gave D-glucose and geniposidic acid. The ¹H- and ¹³C-NMR (*Table*), ¹H,¹H-COSY (*Fig. 2*), HMQC, HMBC (*Fig. 2*), and NOESY data allowed to assign the structure and the absolute configuration of **1** as shown in *Fig. 1*. Daphmacropodosidine A (**1**) is the first example of an iridoid alkaloid in which the alkaloid moiety is located at C(3') of the β-D-glucose moiety instead of C(6') in all iridoid alkaloids reported.

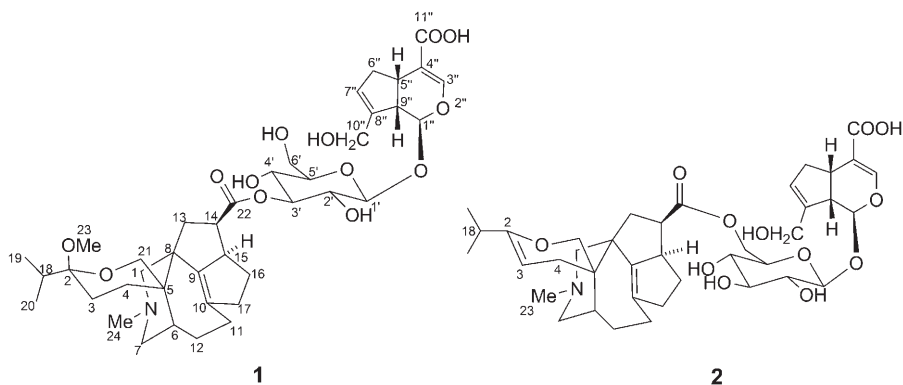


Fig. 1. *Daphmacropodosidine A (1) and B (2)*. Trivial atom numbering.

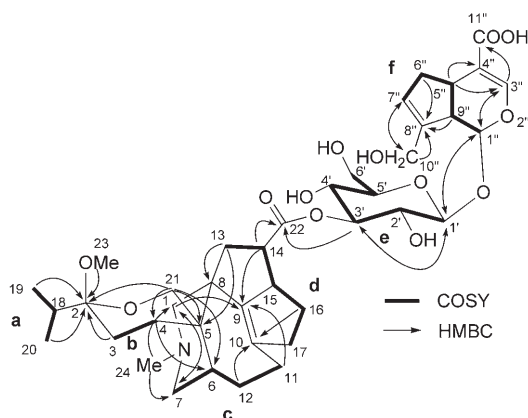


Fig. 2. *Selected 2D-NMR correlations for Daphmacropodosidine A (1)*

The ^{13}C -NMR spectrum of **1** showed 40 C-signals, attributed to 9 quaternary C-atoms (6 sp^2 , 3 sp^3) and 14 CH (2 sp^2 , 12 sp^3), 13 CH_2 (sp^3), and 4 Me (including 1 MeO and 1 MeN moieties). Among the 13 degrees of unsaturation, 2 were assigned to the O-bearing $\text{C}=\text{O}$ ($\delta(\text{C})$ 175.9 and 175.7) and 3 to $\text{C}=\text{C}$ bonds ($\delta(\text{C})$ 118.5 and 148.7, $\delta(\text{C})$ 128.4 and 145.0, and $\delta(\text{C})$ 135.4 and 146.5), thus **1** was inferred to possess 8 rings. Analysis of the ^1H , ^1H -COSY (Fig. 2) and HMQC data of **1** led to the identification of the six partial structures **a** (C(18) to C(20)), **b** (C(3)–C(4)), **c** (C(6)–C(7) and C(11)–C(12)), **d** (C(13) to C(17)), **e** (C(1') to C(6')), and **f** (C(5'') to C(7'') and C(9'')) as well as three isolated CH_2 , *i.e.*, CH_2 (1), CH_2 (21), and CH_2 (10'')¹. Further analysis of the HMBC plot allowed to attribute these six partial structures and three isolated CH_2 to three substructures: an alkaloid moiety, a glucose moiety, and a de-*O*-methylgenipin moiety, which were consistent with those of daphcalycinosidine B [5]. However, comparison of the ^1H - and ^{13}C -NMR spectra of **1** with those of daphcalycinosidine B revealed differences. In the ^1H -NMR spectrum, the signals of H–C(2'), H–C(3'), and H–C(4') were shifted downfield from $\delta(\text{H})$ 3.23, 3.40, and 3.30 in daphcalycinosidine B to $\delta(\text{H})$ 3.35, 4.95, and 3.46 in **1**, and the signals of H–C(5'), H_a–C(6'), and H_b–C(6') were shifted upfield from $\delta(\text{H})$ 3.48, 4.48, and 4.14 in daphcalycinosidine B to $\delta(\text{H})$ 3.32, 3.84, and 3.66 in **1**. Furthermore, in the ^{13}C -NMR spectra, the signals

¹) Trivial atom numbering; for systematic names, see *Exper. Part*.

Table. ¹H- and ¹³C-NMR Data of Compounds **1** and **2**. δ in ppm, J in Hz.

	1 (CD ₃ OD)		2 (C ₅ D ₅ N)	
	δ(H)	δ(C)	δ(H)	δ(C)
CH ₂ (1)	2.55, 2.43 (each <i>d</i> , <i>J</i> = 10.8)	61.2 (<i>t</i>)	2.32, 2.01 (each <i>d</i> , <i>J</i> = 10.4)	64.4 (<i>t</i>)
C(2)		102.3 (<i>s</i>)		159.6 (<i>s</i>)
CH ₂ (3) or H–C(3)	1.73, 1.35 (<i>2m</i>)	22.8 (<i>t</i>)	4.45 (br. <i>d</i> , <i>J</i> = 4.2)	91.2 (<i>d</i>)
CH ₂ (4)	1.96, 1.58 (<i>2m</i>)	23.2 (<i>t</i>)	2.26 (<i>dd</i> , <i>J</i> = 16.8, 4.2), 1.88 (<i>d</i> , <i>J</i> = 16.8)	27.6 (<i>t</i>)
C(5)		37.7 (<i>s</i>)		37.2 (<i>s</i>)
H–C(6)	2.31 (<i>m</i>)	34.1 (<i>d</i>)	2.09 (<i>m</i>)	34.8 (<i>d</i>)
CH ₂ (7)	2.90 (<i>d</i> , <i>J</i> = 12.4), 2.77 (overlap)	56.7 (<i>t</i>)	2.63 (<i>d</i> , <i>J</i> = 11.6), 2.46 (<i>m</i>)	55.5 (<i>t</i>)
C(8)		47.3 (<i>s</i>)		46.8 (<i>s</i>)
C(9)		146.5 (<i>s</i>)		147.3 (<i>s</i>)
C(10)		135.4 (<i>s</i>)		133.5 (<i>s</i>)
CH ₂ (11)	2.11, 1.65 (<i>2m</i>)	27.8 (<i>t</i>)	2.64, 2.40 (<i>2m</i>)	27.6 (<i>t</i>)
CH ₂ (12)	2.45, 2.27 (<i>2m</i>)	27.8 (<i>t</i>)	1.97, 1.62 (<i>2m</i>)	27.7 (<i>t</i>)
CH ₂ (13)	2.76, 1.68 (<i>2m</i>)	40.5 (<i>t</i>)	2.47, 1.54 (<i>2m</i>)	40.0 (<i>t</i>)
H–C(14)	3.00 (<i>m</i>)	44.2 (<i>d</i>)	2.95 (<i>m</i>)	49.9 (<i>d</i>)
H–C(15)	3.55 (<i>m</i>)	56.0 (<i>d</i>)	3.53 (<i>m</i>)	55.5 (<i>d</i>)
CH ₂ (16)	1.83, 1.73 (<i>2m</i>)	29.0 (<i>t</i>)	1.96, 1.56 (<i>2m</i>)	28.9 (<i>t</i>)
CH ₂ (17)	2.57, 2.33 (<i>2m</i>)	43.6 (<i>t</i>)	2.48, 2.36 (<i>2m</i>)	43.0 (<i>t</i>)
H–C(18)	2.04 (<i>m</i>)	32.5 (<i>d</i>)	2.33 (<i>m</i>)	32.6 (<i>d</i>)
Me(19)	0.84 (<i>d</i> , <i>J</i> = 6.9)	16.7 (<i>q</i>)	1.08 (<i>d</i> , <i>J</i> = 6.8)	20.7 (<i>q</i>)
Me(20)	0.93 (<i>d</i> , <i>J</i> = 6.7)	17.7 (<i>q</i>)	1.08 (<i>d</i> , <i>J</i> = 6.8)	20.8 (<i>q</i>)
CH ₂ (21)	4.06, 3.88 (each <i>d</i> , <i>J</i> = 12.4)	64.3 (<i>t</i>)	4.72, 4.38 (each <i>d</i> , <i>J</i> = 11.6)	70.1 (<i>t</i>)
C(22)		175.9 (<i>s</i>)		175.3 (<i>s</i>)
Me(23)O or Me(23)N	3.17 (<i>s</i>)	47.2 (<i>q</i>)	2.11 (<i>s</i>)	46.6 (<i>q</i>)
Me(24)N	2.34 (<i>s</i>)	46.8 (<i>q</i>)		
H–C(1')	4.77 (<i>d</i> , <i>J</i> = 7.6)	100.2 (<i>d</i>)	5.39 (<i>d</i> , <i>J</i> = 7.8)	100.7 (<i>d</i>)
H–C(2')	3.35 (<i>m</i>)	73.2 (<i>d</i>)	4.06 (<i>m</i>)	74.9 (<i>d</i>)
H–C(3')	4.95 (<i>t</i> , <i>J</i> = 7.0)	78.4 (<i>q</i>)	4.25 (<i>t</i> , <i>J</i> = 8.9)	78.2 (<i>d</i>)
H–C(4')	3.46 (<i>m</i>)	69.7 (<i>d</i>)	3.99 (<i>t</i> , <i>J</i> = 8.9)	71.6 (<i>d</i>)
H–C(5')	3.32 (<i>m</i>)	78.3 (<i>q</i>)	4.09 (<i>m</i>)	75.8 (<i>d</i>)
CH ₂ (6')	3.84 (<i>d</i> , <i>J</i> = 12.0), 3.66 (<i>dd</i> , <i>J</i> = 12.0, 5.1)	62.2 (<i>t</i>)	5.01 (<i>d</i> , <i>J</i> = 11.3), 4.55 (<i>dd</i> , <i>J</i> = 11.3, 6.7)	64.6 (<i>t</i>)
H–C(1'')	5.05 (<i>d</i> , <i>J</i> = 7.5)	97.8 (<i>d</i>)	5.68 (<i>d</i> , <i>J</i> = 7.3)	97.7 (<i>d</i>)
H–C(3'')	7.22 (<i>s</i>)	148.7 (<i>d</i>)	7.94 (<i>s</i>)	151.9 (<i>d</i>)
C(4'')		118.5 (<i>s</i>)		113.2 (<i>s</i>)
H–C(5'')	3.21 (<i>m</i>)	37.7 (<i>d</i>)	3.57 (<i>m</i>)	36.2 (<i>d</i>)
CH ₂ (6'')	2.83, 2.06 (<i>2m</i>)	40.0 (<i>t</i>)	3.15, 2.44 (<i>2m</i>)	39.6 (<i>t</i>)
H–C(7'')	5.76 (<i>s</i>)	128.4 (<i>d</i>)	6.02 (<i>s</i>)	127.2 (<i>d</i>)
C(8'')		145.0 (<i>s</i>)		145.6 (<i>s</i>)
H–C(9'')	2.63 (<i>m</i>)	47.5 (<i>d</i>)	3.13 (<i>m</i>)	47.0 (<i>d</i>)
CH ₂ (10'')	4.27, 4.16 (each <i>d</i> , <i>J</i> = 13.2)	61.6 (<i>t</i>)	4.77, 4.65 (each <i>d</i> , <i>J</i> = 14.2)	61.0 (<i>t</i>)
C(11'')		175.7 (<i>s</i>)		169.8 (<i>s</i>)

of C(3') and C(5') were shifted downfield from $\delta(\text{C})$ 77.6 and 75.8 in daphcalycinosidine B to $\delta(\text{C})$ 78.4 and 78.3 in **1**, while the signals of C(2'), C(4'), and C(6') were shifted upfield from $\delta(\text{C})$ 74.8, 71.6, and 64.5 in daphcalycinosidine B to $\delta(\text{C})$ 73.2, 69.7, and 62.2 in **1**. These differences suggested that **1** and daphcalycinosidine B were connected at different positions of the alkaloid and the glucose moieties. Moreover, in the HMBC of **1**, no correlation $\text{CH}_2(6')/\text{C}(22)$ was observed, indicating that the alkaloid moiety was not located at C(6'), while the presence of the strong correlation of H–C(3') at $\delta(\text{H})$ 4.95 with C(22) and anomeric C(1') established the connection of the alkaloid moiety to C(3') in **1**. In the NOESY plot, the strong correlations H–C(14)/H–C(15) and H–C(5'')/H–C(9'') indicated that **1** possessed the same relative configuration as daphcalycinosidine B.

Daphmacropodosidine B (**2**) was isolated as optically active white amorphous powder exhibiting UV absorptions at 308.6 (2.52) and 377.8 (2.26) nm. The HR-ESI-MS ($[M + \text{H}]^+$ at m/z 728.3626) suggested a molecular formula $\text{C}_{39}\text{H}_{53}\text{NO}_{12}$ with 14 degrees of unsaturation. Extensive analysis of the ^1H - and ^{13}C -NMR (Table), ^1H , ^1H -COSY, HMBC, and NOESY data as well as their comparison with those of caldaphnidine F [6c], elucidated **2** as a dehydration derivative of caldaphnidine F.

The ^{13}C -NMR signals for C(2) and C(3) were shifted downfield from $\delta(\text{C})$ 97.8 and 26.0 in caldaphnidine F to $\delta(\text{C})$ 159.6 and 91.2 in **2**, which indicated the presence of a C=C bond between C(2) and C(3) in **2**. Furthermore the chemical shift of H–C(3) at $\delta(\text{H})$ 4.45 was typical for an olefinic proton, and in addition, the HMQC data showed that the C(3) signal was arising from a methine moiety. The correlations H–C(3)/ $\text{CH}_2(4)$ in the ^1H , ^1H COSY plot and H–C(3)/C(2), C(3), C(4), C(5), and C(18) in the HMBC plot supported this assignment.

Two known iridoid alkaloids were identified as daphcalycinosidine B [5] and caldaphnidine F [6c] by comparing the spectral data with those reported in the literature and with those of authentic samples.

Experimental Part

General. Optical rotations: *Jasco-DIP-370* digital polarimeter. M.p.: *YuHua-X-4* apparatus. IR Spectra: *Bio-Rad FTS-135* infrared spectrometer; KBr pellets; in cm^{-1} . UV Spectra: *UV-210A* spectrometer; λ_{max} (log ϵ) in nm. 1D and 2D-NMR Spectra: *DRX-500* spectrometers; δ in ppm, J in Hz; Me_4Si as internal standard, measured in CD_3OD or $\text{C}_5\text{D}_5\text{N}$. HPLC: *Agilent-1100* series; *Waters Spherisorb® NH₂* (5 μm , 4.6×250 mm) anal. column; *LiChroprep® Rp-18* (43–63 μm ; *Merck*). Column chromatography (CC): silica gel (200–300 mesh; *Qindao Marine Chemical Factory*, Qindao, China).

Plant Material. The fresh fruits of *D. macropodum* were collected in Lushan of Jiangxi Province, P. R. China, in November 2005, and identified by Prof. *Xun Gong* of the Kunming Institute of Botany, CAS.

Extraction and Isolation. The fresh fruits of *D. macropodum* were extracted with 95% EtOH (3×40 l) for 4, 3, and 2 h at refluxing temp. The combined extract was concentrated, the residue suspended in H_2O , and after acidification with tartaric acid to pH 3, extracted with CHCl_3 . The H_2O -soluble fraction was basified to pH 9 with 10% aq. NH_3 soln. and extracted with CHCl_3 and BuOH. The BuOH fraction was concentrated and afforded a mixture of H_2O -soluble alkaloids (53 g), which was purified by CC (silica gel, $\text{CHCl}_3/\text{MeOH}$ 99:1 \rightarrow 1:1, then MeOH): *Fractions A–E*. *Fr. C* (12.8 g) was subjected to prep. HPLC on *RP-18*, 0 \rightarrow 40% MeOH/ H_2O): *Fr. C1–C5*. *Fr. C3* (2.1 g) was divided into *Fr. D1–D4* by CC (silica gel, AcOEt/MeOH 9:1 \rightarrow 6:4, then MeOH). Daphmacropodosidine A (14 mg, **1**) was obtained from *Fr. D3* (33 mg) by prep. TLC ($\text{CHCl}_3/\text{MeOH}$ 4:1). Daphmacropodosidine B (8 mg; **2**), daphcalycinosidine B (27 mg), and caldaphnidine F (16 mg) were isolated from 1/12 of *Fr. C4* (300 mg) by anal. HPLC (MeCN/ H_2O 2:8).

Daphmacropodosidine A (= 1*S*,4*aS*,7*aS*)-1- $\{3\text{-O-}\{[(3'S,4'S,6'S,8*a*R,9R,10*a*R)-2,3,4,5,5',6,6',7,8,8*a*,9,10\text{-Dodecahydro-6'-methoxy-2-methyl-6'-(1-methylethyl)spiro[1*H*-4,10*a*-methanopentaleno[1,6-*cd*]-$

azonine-11,3'-(4H)-[2H]pyran]-9-yl]carbonyl]- β -D-glucopyranosyl]oxy]-1,4a,5,7a-tetrahydro-7-(hydroxymethyl)cyclopenta[c]pyran-4-carboxylic Acid; **1**): Colorless crystals. M.p. 176–178° (MeOH). $[\alpha]_D^{29.3} = -43.5$ ($c = 0.84$, MeOH). UV (MeOH): 307.2 (2.73), 374.4 (2.42). IR (KBr): 3422, 2930, 1729, 1643, 1549, 1458, 1400, 1158, 1082, 1042. ^1H - and ^{13}C -NMR: Table. HR-ESI-MS: 760.3910 ($[M + H]^+$, $\text{C}_{40}\text{H}_{58}\text{NO}_{13}^+$; calc. 760.3908).

Daphmacropodosidine B (= (1S,4aS,7aS)-1-[[6-O-[[[(3'S,4S,8aR,9R,10aR)-2,3,4,5,6,7,8,8a,9,10-Decahydro-2-methyl-6'-(1-methylethyl)spiro[1H-4,10a-methanopentaleno[1,6-cd]azonine-11,3'-(4H)-[2H]pyran]-9-yl]carbonyl]- β -D-glucopyranosyl]oxy]-1,4a,5,7-tetrahydro-7-(hydroxymethyl)cyclopenta[c]pyran-4-carboxylic Acid; **2**): White amorphous powder. M.p. 196–200° (MeOH). $[\alpha]_D^{29.3} = 0.0$ ($c = 0.26$, MeOH). UV (MeOH): 308.6 (2.52), 377.8 (2.26). IR (KBr): 3425, 2927, 1735, 1641, 1549, 1452, 1397, 1169, 1146, 1086, 1044. ^1H - and ^{13}C -NMR: Table. HR-ESI-MS: 728.3626 ($[M + H]^+$, $\text{C}_{39}\text{H}_{54}\text{NO}_{12}^+$; calc. 728.3646).

Hydrolysis of Daphmacropodosidine A (**1**) and Daphmacropodosidine B (**2**). A soln. of **1** (1 mg) in 1% KOH soln. (1 ml) was stirred at r.t. for 4 h. Then the mixture was acidified with 1M HCl. D-Glucose and geniposidic acid were identified by co-TLC with authentic samples.

Hydrolysis of **2** (1 mg) by the same procedure afforded also D-glucose and geniposidic acid.

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